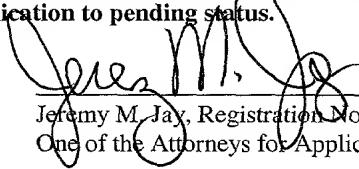
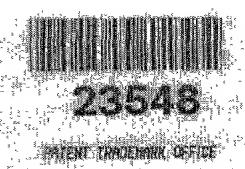


JOHNSON & JOHNSON TO 24 AUG 2001

U.S. DEPARTMENT OF COMMERCE PATENT AND TRADEMARK OFFICE		ATTORNEY DOCKET NO.
<b>TRANSMITTAL LETTER TO THE UNITED STATES DESIGNATED/ELECTED OFFICE (DO/EO/US) CONCERNING A FILING UNDER 35 USC 371</b>		440508/PALL
INTERNATIONAL APPLICATION NO. PCT/US00/04745	INTERNATIONAL FILING DATE February 25, 2000	U.S. APPLICATION NO. <b>09/914165</b>
TITLE OF INVENTION Negatively Charged Membrane		PRIORITY DATE CLAIMED February 25, 1999
APPLICANT(S) FOR DO/EO/US HOU ET AL.		
Applicant herewith submits to the United States Designated/Elected Office (DO/EO/US) the following items and other information:		
1. <input checked="" type="checkbox"/> This is a <b>FIRST</b> submission of items concerning a filing under 35 USC 371.		
2. <input type="checkbox"/> This is a <b>SECOND</b> or <b>SUBSEQUENT</b> submission of items concerning a filing under 35 USC 371.		
3. <input checked="" type="checkbox"/> This is an express request to begin national examination procedures (35 USC 371(f)).		
4. <input checked="" type="checkbox"/> The US has been elected by the expiration of 19 months from the priority date (PCT Article 31).		
5. <input checked="" type="checkbox"/> A copy of the International Application as filed (35 USC 371(c)(2)) a. <input type="checkbox"/> is attached hereto (required only if not communicated by the International Bureau). b. <input type="checkbox"/> has been communicated by the International Bureau. c. <input checked="" type="checkbox"/> is not required, as the application was filed in the United States Receiving Office (RO/US).		
6. <input type="checkbox"/> An English language translation of the International Application as filed (35 USC 371(c)(2)).		
7. <input checked="" type="checkbox"/> Amendments to the claims of the International Application under PCT Article 19 (35 USC 371(c)(3)) a. <input type="checkbox"/> are attached hereto (required only if not communicated by the International Bureau). b. <input type="checkbox"/> have been communicated by the International Bureau. c. <input type="checkbox"/> have not been made; however, the time limit for making such amendments has NOT expired. d. <input checked="" type="checkbox"/> have not been made and will not be made.		
8. <input type="checkbox"/> An English language translation of the amendments to the claims under PCT Article 19 (35 USC 371(c)(3)).		
9. <input type="checkbox"/> An oath or declaration of the inventor(s) (35 USC 371(c)(4)).		
10. <input checked="" type="checkbox"/> An English language translation of the annexes to the International Preliminary Examination Report under PCT Article 36 (35 USC 371(c)(5)).		
11. Nucleotide and/or Amino Acid Sequence Submission a. <input type="checkbox"/> Computer Readable Form (CRF) b. Specification Sequence Listing on: i. <input type="checkbox"/> CD-ROM or CD-R (2 copies); or ii. <input type="checkbox"/> Paper Copy c. <input type="checkbox"/> Statement verifying identity of above copies		
<b>Items 12 to 19 below concern other document(s) or information included:</b>		
12. <input type="checkbox"/> An Information Disclosure Statement under 37 CFR 1.97 and 1.98. <input type="checkbox"/> Form PTO-1449 <input type="checkbox"/> Copies of Listed Documents		
13. <input type="checkbox"/> An assignment for recording. A separate cover sheet in compliance with 37 CFR 3.28 and 3.31 is included.		
14. <input checked="" type="checkbox"/> A <b>FIRST</b> preliminary amendment. <input type="checkbox"/> A <b>SECOND</b> or <b>SUBSEQUENT</b> preliminary amendment.		
15. <input type="checkbox"/> A substitute specification.		
16. <input type="checkbox"/> A change of power of attorney and/or address letter.		
17. <input checked="" type="checkbox"/> Application Data Sheet Under 37 CFR 1.76		
18. <input checked="" type="checkbox"/> Return Receipt Postcard		
19. <input checked="" type="checkbox"/> Other items or information: An International Search Report, Form PCT/ISA/210; An International Preliminary Examination Report, Form PCT/IPEA/409 with Annexes		

U.S. APPLICATION NO.	<b>09/914165</b>	INTERNATIONAL APPLICATION NO.	ATTORNEY DOCKET NO. 440508/PALL																					
20. <input checked="" type="checkbox"/> The following fees are submitted:			CALCULATIONS	PTO USE ONLY																				
<b>Basic National Fee (37 CFR 1.492(a)(1)-(5)):</b>																								
Neither international preliminary examination fee (37 CFR 1.482) nor international search fee (37 CFR 1.445(a)(2)) paid to USPTO and International Search Report not prepared by the EPO or JPO..... \$1,000.00																								
International preliminary examination fee (37 CFR 1.482) not paid to USPTO but International Search Report prepared by the EPO or JPO..... \$ 860.00																								
International preliminary examination fee (37 CFR 1.482) paid to USPTO, but international search fee (37 CFR 1.445(a)(2)) paid to USPTO..... \$ 710.00																								
International preliminary examination fee paid to USPTO (37 CFR 1.482) but all claims did not satisfy provisions of PCT Article 33(1)-(4) ..... \$ 690.00																								
International preliminary examination fee paid to USPTO (37 CFR 1.482) and all claims satisfied provisions of PCT Article 33(1) to (4)..... \$ 100.00																								
<b>ENTER APPROPRIATE BASIC FEE AMOUNT=</b>			\$860.00																					
Surcharge of \$130.00 for furnishing the National fee or oath or declaration later than <input type="checkbox"/> 20 <input type="checkbox"/> 30 months from the earliest claimed priority date			\$																					
<table border="1" style="width: 100%; border-collapse: collapse;"> <thead> <tr> <th>CLAIMS</th> <th>NUMBER FILED</th> <th>NUMBER EXTRA</th> <th>RATE</th> <th></th> </tr> </thead> <tbody> <tr> <td>Total Claims</td> <td>48</td> <td>-20=</td> <td>28</td> <td>x \$ 18.00 \$504.00</td> </tr> <tr> <td>Independent Claims</td> <td>5</td> <td>- 3 =</td> <td>2</td> <td>x \$ 80.00 \$160.00</td> </tr> <tr> <td colspan="3"><input checked="" type="checkbox"/> Multiple Dependent Claim(s) (if applicable)</td> <td>+\$270.00</td> <td>\$</td> </tr> </tbody> </table>			CLAIMS	NUMBER FILED	NUMBER EXTRA	RATE		Total Claims	48	-20=	28	x \$ 18.00 \$504.00	Independent Claims	5	- 3 =	2	x \$ 80.00 \$160.00	<input checked="" type="checkbox"/> Multiple Dependent Claim(s) (if applicable)			+\$270.00	\$		
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<input checked="" type="checkbox"/> Multiple Dependent Claim(s) (if applicable)			+\$270.00	\$																				
<b>TOTAL OF ABOVE CALCULATIONS=</b>			\$																					
<input checked="" type="checkbox"/> Applicant claims small entity status. See 37 CFR 1.27. The fees indicated above are reduced by 1/2.			\$																					
<b>SUBTOTAL=</b>			\$1,524.00																					
Processing fee of \$130.00 for furnishing English Translation later than <input type="checkbox"/> 20 <input type="checkbox"/> 30 months from the earliest claimed priority date.			\$																					
<b>TOTAL NATIONAL FEE=</b>			\$1,524.00																					
Fee for recording the enclosed assignment. The assignment must be accompanied by an appropriate cover sheet. \$40.00 per property			+ \$																					
<b>TOTAL FEE ENCLOSED=</b>			\$1,524.00																					
			Amount to be: refunded \$																					
			charged: \$																					
<p>a. <input checked="" type="checkbox"/> A check in the amount of \$1,524.00 to cover the above fee is enclosed.</p> <p>b. <input type="checkbox"/> Please charge Deposit Account No. 12-1216 in the amount of \$ to cover the above fees. A duplicate copy of this sheet is enclosed.</p> <p>c. <input checked="" type="checkbox"/> The Commissioner is hereby authorized to charge any additional fees which may be required, or credit any overpayment to Deposit Account No. 12-1216. A duplicate copy of this sheet is enclosed.</p>																								
<p><b>NOTE: Where an appropriate time limit under 37 CFR 1.494 or 1.495 has not been met, a petition to revive (37 CFR 1.137(a) or (b)) must be filed and granted to restore the application to pending status.</b></p>																								
<p>SEND ALL CORRESPONDENCE TO:</p> <p style="text-align: center;">          Jeremy M. Jay, Registration No. 33,587          One of the Attorneys for Applicant(s)</p> <p style="text-align: center;">24 Aug. 2001</p> <p>Date</p>																								
<p>  <b>23548</b>  <small>PATENT TRADEMARK OFFICE</small></p>																								

PATENT  
Attorney Docket No. 440508/PALL

**IN THE UNITED STATES PATENT AND TRADEMARK OFFICE**

In re Application of:

HOU et al.

Application No. Unassigned

Art Unit: Unassigned

Filed: Herewith

Examiner: Unassigned

For: NEGATIVELY CHARGED MEMBRANE

**PRELIMINARY AMENDMENT**

Commissioner for Patents  
Washington, D.C. 20231

Dear Sir:

Prior to the examination of the above-identified patent application, please enter the following amendments and consider the following remarks. Pages showing the requested changes are attached. The amendments indicated herein apply to the claims present at the conclusion of the international phase of the PCT application.

*IN THE SPECIFICATION:*

*Replace paragraph 1 with:*

This application is a 35 U.S.C. § 371 of PCT/US00/04745, filed on February 25, 2000, which claims priority of U.S. Provisional Patent Application No. 60/121,668, filed on February 25, 1999, the disclosure of which is incorporated herein by reference in its entirety.

*IN THE CLAIMS:*

*Please replace the indicated claims with:*

3. (Amended) The negatively charged microporous membrane of claim 1, wherein the N-(hydroxyalkyl)- or N-(alkoxyalkyl)- acrylamide includes an alkyl group of 4 carbon atoms or less.

4. (Amended) The negatively charged microporous membrane of claim 1, wherein the crosslinked coating includes a hydroxyl-rich material.

6. (Amended) The negatively charged microporous membrane of claim 1, wherein said negatively charged group is a sulfonic or carboxylic acid.

18. (Amended) The negatively charged microporous membrane of claim 1, wherein said polymer includes an N-(alkoxymethyl) acrylamide.
21. (Amended) The negatively charged microporous membrane of claim 1 having a dynamic protein binding capacity of about 25 mg/ml lysozyme or more.
23. (Amended) The negatively charged microporous membrane of claim 1, wherein said porous substrate comprises a substrate polymer.
26. (Amended) The negatively charged microporous membrane of claim 1, wherein said porous substrate is hydrophilic.
29. (Amended) The process of claim 27, wherein said negatively charged group is a sulfonic or carboxylic acid.
30. (Amended) The process of claim 27, wherein said unsaturated monomer having a negatively charged group is an acrylic monomer having a sulfonic or carboxylic acid group.
33. (Amended) The process of claim 27, wherein said porous substrate comprises a substrate polymer.
34. (Amended) The negatively charged microporous membrane prepared by the process of claim 27.
35. (Amended) A device comprising the negatively charged microporous membrane of claim 1.
36. (Amended) A process for separating positively charged material from a fluid, said process comprising placing said fluid in contact with the negatively charged microporous membrane of claim 1, so as to adsorb the positively charged material to said membrane.
38. (Amended) A process for transferring biomolecules from an electrophoresis gel comprising contacting said electrophoresis gel with a membrane of claim 1 and transferring the biomolecules to the membrane.

40. (Amended) The process of claim 38, further including recovering the positively charged material adsorbed on the membrane.

Please add the following claims:

41. (New) A negatively charged microporous membrane comprising a porous substrate and a crosslinked coating comprising negatively charged groups and amide-amide and amide-ester crosslinks, wherein the amide-amide crosslink has the formula -C(=O)NH-R-NH C(=O)- and the amide-ester crosslink has the formula -C(=O)O-R-NH-C(=O)-, wherein R is a divalent radical.

42. (New) The negatively charged microporous membrane of claim 41, wherein the divalent radical is an alkoxyalkyl radical.

43. (New) The negatively charged microporous membrane of claim 42, wherein the alkoxyalkyl radical is -CH<sub>2</sub>-O-CH<sub>2</sub>-.

44. (New) The negatively charged microporous membrane of claim 42, wherein the alkoxyalkyl radical is -CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-O-CH<sub>2</sub>-.

45. (New) The negatively charged microporous membrane of claim 41, wherein the crosslinked coating comprises a polymerized unsaturated monomer having a negatively charged group and a polymerized hydrophilic non-ionic unsaturated monomer.

46. (New) The negatively charged microporous membrane of claim 45, wherein the hydrophilic non-ionic unsaturated monomer is an acrylic monomer.

47. (New) The negatively charged microporous membrane of claim 45, wherein the monomer having a negatively charged group is an acrylic monomer.

48. (New) The negatively charged microporous membrane of claim 41, wherein the negatively charged group is a sulfonic acid group or carboxylic acid group.

**PATENT**  
Attorney Docket No. 440508/PALL

**IN THE UNITED STATES PATENT AND TRADEMARK OFFICE**

In re Application of:

HOU et al.

Application No. Unassigned

Filed: Herewith

For: NEGATIVELY CHARGED  
MEMBRANE

Art Unit: Unassigned

Examiner: Unassigned

**AMENDMENTS TO THE SPECIFICATION AND  
CLAIMS MADE VIA PRELIMINARY AMENDMENT**

*Amendments to paragraph 1:*

This application is a 35 U.S.C. § 371 of PCT/US00/04745, filed on February 25, 2000,  
which This application claims priority of U.S. Provisional Patent Application No. 60/128,668  
60/121,668, filed on February 25, 1999, the disclosure of which is incorporated herein by  
reference in its entirety.

*Amendments to existing claims:*

3. (Amended) The negatively charged microporous membrane of ~~claim 1 or 2,~~ wherein the N-(hydroxyalkyl)- or N-(alkoxyalkyl)- acrylamide includes an alkyl group of 4 carbon atoms or less.

4. (Amended) The negatively charged microporous membrane of ~~any of claims 1-3, claim 1,~~ wherein the crosslinked coating includes a hydroxyl-rich material.

6. (Amended) The negatively charged microporous membrane of ~~any of claims 1-5, claim 1,~~ wherein said negatively charged group is a sulfonic or carboxylic acid.

18. (Amended) The negatively charged microporous membrane of ~~any of claims 1-17, claim 1,~~ wherein said polymer includes an N-(alkoxymethyl) acrylamide.

21. (Amended) The negatively charged microporous membrane of ~~any of claims 1-20 claim 1~~ having a dynamic protein binding capacity of about 25 mg/ml lysozyme or more.

23. (Amended) The negatively charged microporous membrane of ~~any of claims 1-22, claim 1,~~ wherein said porous substrate comprises a substrate polymer.

26. (Amended) The negatively charged microporous membrane of ~~any of claims 1-22, claim 1,~~ wherein said porous substrate is hydrophilic.

29. (Amended) The process of claim 27~~or 28~~, wherein said negatively charged group is a sulfonic or carboxylic acid.

30. (Amended) The process of ~~any of claims 27-29, claim 27~~, wherein said unsaturated monomer having a negatively charged group is an acrylic monomer having a sulfonic or carboxylic acid group.

33. (Amended) The process of ~~any of claims 27-32, claim 27~~, wherein said porous substrate comprises a substrate polymer.

34. (Amended) The negatively charged microporous membrane prepared by the process of ~~any of claims 27-33 claim 27~~.

35. (Amended) A device comprising the negatively charged microporous membrane of ~~any of claims 1-26 and 34 claim 1~~.

36. (Amended) A process for separating positively charged material from a fluid, said process comprising placing said fluid in contact with the negatively charged microporous membrane of ~~any of claims 1-26 and 34 claim 1~~, so as to adsorb the positively charged material to said membrane.

38. (Amended) A process for transferring biomolecules from an electrophoresis gel comprising contacting said electrophoresis gel with a membrane of ~~any of claims 1-27 and 34 claim 1~~ and transferring the biomolecules to the membrane.

40. (Amended) The process of claim 38~~or 39~~, further including recovering the positively charged material adsorbed on the membrane.

PATENT  
Attorney Docket No. 440508/PALL

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

In re Application of:

HOU et al.

Application No. Unassigned

Filed: Herewith

For: NEGATIVELY CHARGED  
MEMBRANE

Art Unit: Unassigned

Examiner: Unassigned

PENDING CLAIMS AFTER ENTRY OF PRELIMINARY AMENDMENT

1. A negatively charged microporous membrane comprising a porous substrate and a crosslinked coating, wherein the crosslinked coating is prepared from a polymer comprising an unsaturated monomer having a negatively charged group, a hydrophilic non-ionic unsaturated monomer, and at least one or more N-(hydroxyalkyl)- or N-(alkoxyalkyl)-acrylamide.
2. The negatively charged microporous membrane of claim 1, wherein the hydrophilic non-ionic unsaturated monomer is an acrylic monomer.
3. The negatively charged microporous membrane of claim 1, wherein the N-(hydroxyalkyl)- or N-(alkoxyalkyl)- acrylamide includes an alkyl group of 4 carbon atoms or less.
4. The negatively charged microporous membrane of claim 1, wherein the crosslinked coating includes a hydroxyl-rich material.
5. The negatively charged microporous membrane of claim 4, wherein the hydroxyl-rich material is a polysaccharide.
6. The negatively charged microporous membrane of claim 1, wherein said negatively charged group is a sulfonic or carboxylic acid.

7. The negatively charged microporous membrane of claim 2, wherein said acrylic monomer is an acrylate or acrylamide.
8. The negatively charged microporous membrane of claim 7, wherein said acrylic monomer is an acrylamide.
9. The negatively charged microporous membrane of claim 8, wherein said acrylamide is an alkylacrylamide.
10. The negatively charged microporous membrane of claim 9, wherein said acrylamide has a sulfonic acid group.
11. The negatively charged microporous membrane of claim 10, wherein said acrylamide is acrylamido-N-alkylsulfonic acid.
12. The negatively charged microporous membrane of claim 9, wherein said alkylacrylamide has a carboxylic acid group.
13. The negatively charged microporous membrane of claim 12, wherein said polymer includes a further acrylic monomer having a carboxylic acid group.
14. The negatively charged microporous membrane of claim 13, wherein said further acrylic monomer is an acrylate.
15. The negatively charged microporous membrane of claim 14, wherein said acrylate is  $\beta$ -carboxyethyl acrylate.
16. The negatively charged microporous membrane of claim 4, wherein said acrylic monomer is a hydroxyacrylic monomer.
17. The negatively charged microporous membrane of claim 16, wherein said hydroxyacrylic monomer is a hydroxyacrylamide or an hydroxyacrylate.

18. The negatively charged microporous membrane of claim 1, wherein said polymer includes an N-(alkoxymethyl) acrylamide.
19. The negatively charged microporous membrane of claim 5, wherein said polysaccharide is dextran.
20. The negatively charged microporous membrane of claim 1, wherein the polymer comprising an unsaturated monomer having a negatively charged group, a hydrophilic non-ionic unsaturated monomer, and at least one or more N-(hydroxyalkyl)- or N-(alkoxyalkyl)-acrylamide is prepared by employing a free radical initiator.
21. The negatively charged microporous membrane of claim 1 having a dynamic protein binding capacity of about 25 mg/ml lysozyme or more.
22. A negatively charged microporous membrane comprising a porous substrate and a crosslinked coating comprising negatively charged groups and amide-amide and amide-ester crosslinks.
23. The negatively charged microporous membrane of claim 1, wherein said porous substrate comprises a substrate polymer.
24. The negatively charged microporous membrane of claim 23, wherein said substrate polymer is selected from the group consisting of polyaromatics, polysulfones, polyolefins, polystyrenes, polyamides, polyimides, cellulose acetates, cellulose nitrates, polycarbonates, polyesters, and fluoropolymers.
25. The negatively charged microporous membrane of claim 24, wherein said substrate polymer is a polysulfone.
26. The negatively charged microporous membrane of claim 1, wherein said porous substrate is hydrophilic.

27. A process for preparing a negatively charged microporous membrane comprising a porous substrate and a crosslinked coating having negatively charged groups, the process comprising:
- (a) providing a porous substrate;
  - (b) contacting said substrate with a polymer comprising an unsaturated monomer having a negatively charged group, a hydrophilic non-ionic unsaturated monomer, and at least one or more of a N-(hydroxyalkyl)- or N-(alkoxyalkyl)- acrylamide;
  - (c) curing the substrate obtained in (b) to obtain the negatively charged microporous membrane; and
  - (d) optionally, extracting the membrane obtained in (c) to remove extractable residue therein.
28. A process for preparing a negatively charged microporous membrane comprising a porous substrate and a crosslinked coating having negatively charged groups, the process comprising:
- (a) providing a porous substrate;
  - (b) contacting said substrate with a polysaccharide and a polymer comprising an unsaturated monomer having a negatively charged group and an N-(hydroxymethyl)- or N-(alkoxymethyl)- acrylamide;
  - (c) curing the substrate obtained in (b) to obtain the negatively charged microporous membrane; and
  - (d) optionally, extracting the membrane obtained in (c) to remove extractable residue therein.
29. The process of claim 27, wherein said negatively charged group is a sulfonic or carboxylic acid.
30. The process of claim 27, wherein said unsaturated monomer having a negatively charged group is an acrylic monomer having a sulfonic or carboxylic acid group.
31. The process of claim 30, wherein said acrylic monomer having a sulfonic or carboxylic acid group is an acrylate or an acrylamide.
32. The process of claim 27, wherein the substrate is contacted in (b) with said polymer and a hydroxyl-rich material.

33. The process of claim 27, wherein said porous substrate comprises a substrate polymer.
34. The negatively charged microporous membrane prepared by the process of claim 27.
35. A device comprising the negatively charged microporous membrane of claim 1.
36. A process for separating positively charged material from a fluid, said process comprising placing said fluid in contact with the negatively charged microporous membrane of claim 1, so as to adsorb the positively charged material to said membrane.
37. The process of claim 36, wherein said positively charged material is a biomolecule.
38. A process for transferring biomolecules from an electrophoresis gel comprising contacting said electrophoresis gel with a membrane of claim 1 and transferring the biomolecules to the membrane.
39. The process of claim 38, wherein said biomolecule is selected from the group consisting of proteins, polypeptides, amino acids, and nucleic acids, and combinations thereof.
40. The process of claim 38, further including recovering the positively charged material adsorbed on the membrane.
41. A negatively charged microporous membrane comprising a porous substrate and a crosslinked coating comprising negatively charged groups and amide-amide and amide-ester crosslinks, wherein the amide-amide crosslink has the formula -C(=O)NH-R-NH C(=O)- and the amide-ester crosslink has the formula -C(=O)O-R-NH-C(=O)-, wherein R is a divalent radical.
42. The negatively charged microporous membrane of claim 41, wherein the divalent radical is an alkoxyalkyl radical.

43. The negatively charged microporous membrane of claim 42, wherein the alkoxyalkyl radical is -CH<sub>2</sub>-O-CH<sub>2</sub>-.
44. The negatively charged microporous membrane of claim 42, wherein the alkoxyalkyl radical is -CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-O-CH<sub>2</sub>-.
45. The negatively charged microporous membrane of claim 41, wherein the crosslinked coating comprises a polymerized unsaturated monomer having a negatively charged group and a polymerized hydrophilic non-ionic unsaturated monomer.
46. The negatively charged microporous membrane of claim 45, wherein the hydrophilic non-ionic unsaturated monomer is an acrylic monomer.
47. The negatively charged microporous membrane of claim 45, wherein the monomer having a negatively charged group is an acrylic monomer.
48. The negatively charged microporous membrane of claim 41, wherein the negatively charged group is a sulfonic acid group or carboxylic acid group.

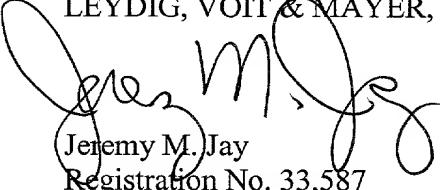
**REMARKS**

The specification has been amended to include a reference to the PCT application, and to indicate that this is the national stage of the PCT application. The specification has also been amended to correct the obvious error in the priority application number, and this amendment merely reflects the rectification made during the international stage. The claims have been amended to remove multiple dependencies. New claims 41-48 have been added and are directed to embodiments of the invention. Claims 1-48 are pending. A complete set of pending claims is attached. No new matter is added by way of this Preliminary Amendment.

If, in the opinion of the Examiner, a telephone conference would expedite the prosecution of the subject application, the Examiner is invited to call the undersigned attorney.

Respectfully submitted,

LEYDIG, VOIT & MAYER, LTD.

  
Jeremy M. Jay  
Registration No. 33,587

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Washington, D.C. 20005  
Telephone: (202) 737-6770  
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Date: 24 Aug. 2001  
JMJ:jj

09/914 165

Rec'd PCT/PTO 24 AUG 2001  
PCT/US00/04745

WO 00/50160

NEGATIVELY CHARGED MEMBRANE

CROSS-REFERENCE TO A RELATED APPLICATION

This application claims priority from U.S. Provisional

- 5 Patent Application No. 60/128,668, filed on February 25, 1999,  
the disclosure of which is incorporated herein by reference in  
its entirety.

FIELD OF THE INVENTION

- 10 The present invention generally relates to negatively charged membranes, and in particular to negatively charged membranes comprising a porous substrate and a crosslinked coating. The membranes find use in the treatment of fluids containing positively charged species such as proteins, e.g.,  
15 in ion-exchange chromatography.

BACKGROUND OF THE INVENTION

- Negatively charged ion-exchange membranes have been proposed for the separation and/or purification of biomolecules such as proteins, amino acids, and nucleic acids. For the ion exchange membrane to perform effectively in the above applications, the membrane should satisfy several important parameters. For example, the membrane should exhibit high rates of fluid flow. The membrane should have high dynamic binding capacity for biomolecules, and should be capable of selectively binding the biomolecules, which have different surface charges. The membrane should, therefore, have low non-specific binding, e.g., resulting from hydrophobic interactions. The membrane should withstand high treatment fluid velocities. The preparation of the membrane should not involve chemistries and processes that are cumbersome to practice. Some of the cation exchange membranes known heretofore suffer from the failure to satisfy one or more of the parameters set forth above.
- Accordingly, there exists a need for a cation exchange membrane that exhibits high rates of fluid flow. There further exists a need for a cation exchange membrane that has

28-02-2001

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high dynamic binding capacity and selectivity for biomolecules. There further exists a need for a membrane that has low non-specific binding or low binding that results from hydrophobic interactions. There further exists a need for a 5 membrane that can withstand high fluid flow velocities. There further exists a need for a membrane that involves preparation chemistries and/or processes that are not cumbersome to practice.

These advantages of the present invention, as well as 10 additional inventive features, will be apparent from the description of the invention provided herein.

U.S. Patent 5,021,160 discloses copolymers synthesized from 2-acrylamido-2-methyl-1-propane sulfonic acid and either N-(isobutoxymethyl) acrylamide or 2-hydroxyethyl methacrylate 15 and a process for preparing anionic charge modified microporous filtration membranes.

WO 98/17377 discloses charged membranes comprising a porous substrate and a cross-linked polyelectrolyte or hydrogel located in the pores of the substrate.

European Patent Application No. 0 474 617 A1 discloses a surface modified support membrane wherein the support membrane has a layer of hydrogel deposited on the surface thereof.

#### BRIEF DESCRIPTION OF THE DRAWINGS

Fig. 1 depicts the breakthrough curve for lysozyme obtained on an embodiment membrane of the present invention. The x-axis represents the filtration time, and the y-axis represents the absorbance of the filtrate at 280 nm and is indicative of the concentration of the protein. See Example 2 30 for additional details.

Fig. 2 depicts the breakthrough curve for lysozyme obtained on another embodiment membrane of the present invention. The x-axis and y-axis are as described in Fig. 1. See Example 3 for additional details.

Fig. 3 depicts the breakthrough curve for lysozyme obtained on another embodiment membrane of the present

28-02-2001

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invention. The x-axis and y-axis are as described in Fig. 1. See Example 4 for additional details.

### BRIEF SUMMARY OF THE INVENTION

- 5 Many of the foregoing needs have been fulfilled by the present invention which provides a negatively charged microporous membrane comprising a porous substrate and a crosslinked coating having negatively charged groups. In a preferred embodiment, the membrane can be prepared from a  
10 polymerized composition comprising an unsaturated monomer having an anionic group, at least one or more N-

(hydroxyalkyl)- and/or N-(alkoxyalkyl)- acrylamides, and a hydrophilic unsaturated monomer.

The present invention further provides a negatively charged microporous membrane comprising a porous substrate and a crosslinked coating prepared from a hydroxyl-rich material such as a polysaccharide and a polymerized composition comprising an unsaturated monomer having an anionic group, at least one or more N-(hydroxymethyl)- and/or N-(alkoxymethyl)- acrylamides, and an initiator.

The present invention further provides a negatively charged microporous membrane having a protein binding capacity of about 25 mg/ml lysozyme or more comprising a porous substrate and a crosslinked coating that provides a fixed negative charge. The present invention further provides a negatively charged microporous membrane comprising a porous substrate and a crosslinked coating comprising a polymer having anionic groups and amide-amide and amide-ester crosslinks.

The membranes of the present invention are advantageously free of covalent bonds or grafts with the substrate.

The present invention further provides a process for preparing an embodiment of the membrane comprising coating a porous substrate with a polymerized composition comprising an anionic group and curing the membrane. The membrane can be optionally washed or leached to remove extractable residue therein.

The present invention further provides devices, e.g., filter devices, chromatographic devices, macromolecular transfer devices, and membrane modules comprising the membranes of the present invention. The present invention further provides a process for separating a positively charged material such as positively charged atoms, molecules, and particulates, and, preferably, biomolecules, from a fluid, the process comprising placing the fluid in contact with the negatively charged microporous membrane so as to adsorb the positively charged material to the membrane. The present invention further provides a process for treating a fluid

containing positively charged materials comprising contacting the fluid with a membrane of the present invention and selectively releasing the positively charged materials. The present invention further provides a process for transferring 5 macromolecules from a device or element such as an electrophoresis gel comprising contacting the gel with the membrane of the present invention and transferring the biomolecules to the membrane.

While the invention has been described and disclosed below 10 in connection with certain preferred embodiments and procedures, it is not intended to limit the invention to those specific embodiments. Rather it is intended to cover all such alternative embodiments and modifications as fall within the spirit and scope of the invention.

15

#### DETAILED DESCRIPTION OF THE PREFERRED EMBODIMENTS

The present invention provides embodiments of negatively charged membranes having high charge density, high water flow rates, high dynamic protein binding capacity, and low non-specific protein binding capacity. The membranes of the 20 present invention find use in cation exchange chromatography and in the separation and/or purification of charged species, especially biomolecules such as proteins.

The present invention provides, in some embodiments, a 25 negatively charged microporous membrane comprising a porous substrate and a crosslinked coating having anionic groups. The crosslinked coating can be prepared from a polymerized composition comprising an unsaturated monomer having an anionic group, at least one or more N-(hydroxyalkyl)- or N-(alkoxyalkyl)- acrylamides, a hydrophilic unsaturated monomer. 30 The composition can further include an initiator. In preferred embodiments, the N-(hydroxyalkyl)- or N-(alkoxyalkyl)- acrylamide is one wherein the alkyl moiety has 4 or less carbon atoms, and more preferably the alkyl moiety 35 is methyl.

In certain embodiments, the membrane comprises a porous substrate and a crosslinked coating prepared from a

polymerized composition comprising an unsaturated monomer having an anionic group, at least one or more N-(hydroxymethyl)- and/or N-(alkoxymethyl)- acrylamides, a hydroxyl-rich material such as a polysaccharide, and

5 optionally an initiator. The present invention further provides a negatively charged microporous membrane comprising a porous substrate and a crosslinked coating comprising a polymer having anionic groups and amide-amide and amide-ester crosslinks.

10 The membrane of the present invention contains fixed anionic groups. The anionic group can be any negatively charged group - sulfonic, carboxylic, phosphonic, and the like, preferably sulfonic or carboxylic acid groups. The coating composition comprises an unsaturated monomer having an anionic group. Any suitable unsaturated monomer - vinyl, vinyl aromatic, acrylic, or other monomer can be used.

The unsaturated monomer preferably is an acrylic monomer. The acrylic monomer can be an acrylate or an acrylamide. The acrylic monomer is preferably an acrylamide. The term "acrylamide" herein refers to unsubstituted as well as substituted monomers having a -C=C-(C=O)-N- moiety. The nitrogen and the C=C carbon atoms can be attached to hydrogen or other nonpolar substituents. An example of such substituents is alkyl. Thus, the substituted acrylamide can be alkylacrylamide. The term "alkyl" herein refers to an alkyl group having from 1 to about 10 carbon atoms, preferably from 1 to about 6 carbon atoms, and more preferably from 1 to about 3 carbon atoms. An example of an acrylamide monomer having a sulfonic acid group is acrylamido-N-alkylsulfonic acid, preferably 2-acrylamido-2-methyl-1-propanesulfonic acid. Preferred examples of acrylic monomers having a carboxylic acid group are 3-acrylamido-3-methylbutanoic acid (AMBA), 2-acrylamidoglycolic acid, and  $\beta$ -carboxyethyl acrylate.

In certain embodiments, the coating composition comprises 35 a hydrophilic unsaturated monomer, e.g., a nonionic hydrophilic unsaturated monomer. Any suitable hydrophilic unsaturated monomer can be used, preferably an acrylic

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monomer. The monomer contains one or more polar groups that contribute hydrophilicity. Examples of such groups include hydroxy, alkoxy, hydroxyalkyl, and amido. Preferred hydrophilic groups are hydroxyl and hydroxyalkyl. Thus,

5 preferred hydrophilic acrylic monomers are hydroxyacrylic and hydroxyalkylacrylic. The acrylic monomer can be an acrylate ester or an acrylamide. An example of a preferred hydroxyalkyl acrylate monomer is hydroxypropyl methacrylate.

The coating composition comprises a crosslinking agent.

10 Any suitable crosslinking agent known to those of ordinary skill in the art can be used. Preferred crosslinking agents include N-(alkoxymethyl)acrylamide and N-(hydroxymethyl)acrylamide. N-(isobutoxymethyl)acrylamide is further preferred.

15 The coating composition preferably comprises an initiator. Any suitable initiator - free radical initiator, photoinitiator, and the like, can be used. A free radical initiator is preferred. An example of a suitable free radical initiator is a persulfate such as ammonium persulfate.

20 Without being bound to any particular theory, it is believed that the use of the three monomers in certain embodiments contributes to increased spatial separation of charges. Thus, it is believed that the distance between the anionic groups is increased. This increased distance  
25 disfavors association of the anionic groups. Accordingly, inter- and/or intra- chain association of anionic groups is reduced compared to a system wherein only an anionic monomer and a crosslinking monomer are employed, particularly in a two monomer system wherein a hydrophilic or hydroxyl-rich material  
30 such as a polysaccharide is not employed. The reduced association makes the negatively charged groups available for interaction with positively charged molecules in the treated fluid. This results, for example, in enhanced dynamic protein binding capacity.

35 The membrane according to some embodiments is made from a coating composition that includes a hydroxyl-rich material, which may be a small molecule or a polymer having a plurality

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of hydroxyl groups, e.g., two, three, four or more hydroxyl groups per molecule. Examples of hydroxyl-rich materials include polysaccharides and polyvinyl alcohol, preferably polysaccharides. Without being bound to any particular mechanism, it is believed that the hydroxyl groups of the hydroxyl-rich material involve in hydrogen bonding with the fluid. The saccharide ring repeat units exert steric effects. Operation of one or both of these mechanisms results in increased charge separation among the anionic groups. The increased charge separation is believed to reduce anion association and facilitate interaction between the anionic sites and the positively charged species in the treated fluid.

Any suitable polysaccharide can be used, preferably a water soluble polysaccharide. An example of a preferred polysaccharide is dextran. The molecular weight of the dextran is below about 40,000,000, e.g., from about 10,000 to about 2,000,000, preferably from about 10,000 to about 500,000, and more preferably from about 10,000 to about 300,000. Particular examples of suitable molecular weights include 110,000 and 148,000.

The coating composition of certain embodiments can be prepared by combining and polymerizing the acrylic monomer having an anionic group, the nonionic hydrophilic monomer, a crosslinking agent, and the initiator. In some embodiments, the coating solution is prepared by combining and polymerizing the acrylic monomer having an anionic group, the polysaccharide, the crosslinking agent, and the initiator.

The polymerization can be carried out in a solvent, preferably in water or water/methanol solution. The polymerization is preferably stopped prior to the formation of a gel or excessive crosslinking. The viscosity of the polymerization solution can be monitored to control the degree of polymerization. The polymerization is carried out for any suitable length of time, e.g., for about 4 hours or more. According to certain embodiments, the polymerization is carried out for a period of from about 4 hours to about 5 hours. According to certain other embodiments, the

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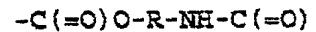
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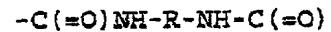
polymerization is carried out for a period of from about 16 hours to about 24 hours. The viscosity of the solution is typically below about 2000 cps (2 Pa.s), e.g., preferably from about 50 cps (0.05 Pa.s) to about 500 cps (0.5 Pa.s), and more 5 preferably from about 100 cps (0.1 Pa.s) to about 500 cps (0.5 Pa.s). According to certain embodiments, the viscosity is from about 100 cps (0.1 Pa.s) to about 250 cps (0.25 Pa.s).

The polymerization solution can contain the anionic acrylic monomer (A), the crosslinking agent (B), and the non- 10 ionic hydrophilic monomer (C) in a suitable ratio. The percentage of each monomer (A, B, or C) can be from about 0.1 to 30% by weight, preferably from about 0.1 to 20% by weight.

It is believed that the crosslinked coating comprises amide-ester crosslinks that form as a result of the reaction 15 of the nonionic hydrophilic monomer with the crosslinking agent. For example, these bonds form as a result of the reaction of the hydroxyl groups in the hydroxyalkyl acrylate with the N-(isobutoxymethyl)- acrylamide. In addition, amide-amide crosslinks also form as a result of the reaction between 20 two N-(isobutoxymethyl)acrylamide monomers. For example, the amide-ester crosslink can have the formula:



wherein R is divalent radical, preferably an alkoxyalkyl radical, and more preferably  $-\text{CH}_2-\text{CH}_2-\text{CH}_2-\text{O}-\text{CH}_2-$ . The amide- 25 amide crosslink can have the formula:



wherein R is divalent radical, preferably an alkoxyalkyl radical, and more preferably  $-\text{CH}_2-\text{O}-\text{CH}_2-$ .

The coating solution contains the anionic polymer 30 prepared as above and, optionally, a polysaccharide, preferably a dextran. The anionic polymer and the polysaccharide can be present in the coating solution in the ratio of from about 100:1 to about 1:100, preferably from about 10:1 to about 1:10, and more preferably from about 5:1 to about 1:5.

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The coating solution contains the anionic polymer and, optionally dextran, in an amount of from about 0.01% to about

15% by weight, preferably from about 0.1% to about 10% by weight, and more preferably from about 0.5% to about 5% by weight of the coating solution. For example, the coating solution can contain 4.5% by weight of polymer and 1.5% by 5 weight of dextran.

The pH of the coating solution can be adjusted suitably. For example, the pH of the coating solution containing a carboxylated polymer can be adjusted to about 3.0 to about 4.0 and preferably about 3.75. The pH of the coating can be 10 adjusted by the addition of an acid or base. An example of a suitable base is 2N NaOH aqueous solution.

The coating solution is coated on a porous substrate, preferably a hydrophilic substrate. The hydrophilic porous substrate can be made of any suitable material; preferably, 15 the substrate comprises a polymer. Examples of suitable polymers include polyaromatics, polysulfones, polyolefins, polystyrenes, polycarbonates, polyamides, polyimides, fluoropolymers, cellulosic polymers such as cellulose acetates and cellulose nitrates, and PEEK. Aromatic polysulfones are preferred. Examples of aromatic polysulfones include 20 polyethersulfone, bisphenol A polysulfone, and polyphenylsulfone. Polyethersulfone is particularly preferred. The porous substrate can have any suitable pore size, for example, a pore size of below about 10  $\mu\text{m}$ , e.g., 25 from about 0.01  $\mu\text{m}$  to about 10  $\mu\text{m}$ , preferably from about 0.1  $\mu\text{m}$  to about 5  $\mu\text{m}$ , and more preferably from about 0.2  $\mu\text{m}$  to about 5  $\mu\text{m}$ . The porous substrate can be asymmetric or, in a preferred embodiment, symmetric.

The porous substrate can be prepared by methods known to 30 those of ordinary skill in the art. For example, the porous substrate can be prepared by a phase inversion process. Thus, a casting solution containing the polymer, a solvent, a pore former, a wetting agent, and optionally a small quantity of a non-solvent is prepared by combining and mixing the 35 ingredients, preferably at an elevated temperature. The resulting solution is filtered to remove any impurities. The casting solution is cast or extruded in the form of a sheet or

hollow fiber. The resulting sheet or fiber is allowed to set or gel as a phase inverted membrane. The set membrane is then leached to remove the solvent and other soluble ingredients.

The porous substrate can be coated with the coating

5 solution by methods known to those of ordinary skill in the art, for example, by dip coating, spray coating, meniscus coating, and the like. Dip coating, for example, can be carried out as follows. The substrate is immersed in the solution for a given period of time sufficient to insure  
10 complete or substantially complete coating of the pore walls. The immersion time can be from about 1 second to 1.0 minute, preferably from about 0.1 minutes to about 0.5 minutes, and more preferably from about 1/6 minute to about 1/3 minute.

15 Following the immersion, the excess coating solution on the substrate is removed by allowing it to drain under gravity or by the use of a squeegee or air knife. The resulting coated substrate is cured to effect the curing or crosslinking of the coating composition.

Thus, the membrane can be cured below 150°C, e.g., at a  
20 temperature of from about 60°C to about 130°C, and preferably at a temperature of from about 80°C to about 130°C, for a suitable period of time, which can range from about 5 minutes to about 120 minutes and preferably from about 5 minutes to about 60 minutes. According to certain embodiments, the  
25 membrane is cured at a temperature of from about 120°C to about 125°C for a period of from about 20 minutes to about 30 minutes.

The resulting membrane can be washed to leach out any extractable in the membrane. Certain embodiments of the  
30 membrane, particularly a membrane having carboxyl functionality, are washed or leached in a basic solution, preferably at a pH of from about 8 to about 12. The leaching liquid can be prepared by adding a base such as sodium hydroxide, sodium carbonate, or sodium bicarbonate. The base  
35 can be added as a solid or a solution. Particular examples of pH's of the leaching liquid are about 11.9, about 11.4, and

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about 8.1. These pH's can be achieved by the use of, e.g., a 2N NaOH solution, sodium carbonate, or sodium bicarbonate.

Illustratively, a carboxylated membrane can be washed or leached at a temperature of from about 37°C to about 93°C or

5 higher and preferably from about 54°C to about 73°C or higher.

A sulfonic acid containing membrane can be washed or leached at a temperature of from about 54°C to about 93°C or higher.

Embodiments of the membrane also can be leached in hot

deionized water, e.g., deionized water held above 73°F. The

10 washing or leaching can be carried out for a suitable length of time, for example, for about 20 to about 30 minutes or more.

According to certain embodiments of the membrane, the washing or leaching can be carried out for about 1 hour or more. The resulting membrane is then dried in air or in an oven to remove the water.

The present invention provides a process for preparing a negatively charged microporous membrane comprising a porous substrate and a crosslinked coating having pendant anionic group. An embodiment of the process comprises:

20 (a) providing a porous substrate;

(b) contacting the substrate with a hydroxyl-rich material and a polymerized composition comprising an unsaturated monomer having an anionic group, at least one or more N-(hydroxyalkyl)- and/or N-(alkoxyalkyl)- acrylamides, a 25 hydrophilic unsaturated monomer, and optionally an initiator;

(c) curing the substrate obtained in (b) to obtain the negatively charged membrane; and

(d) optionally, extracting the membrane obtained in (c) to remove extractable residue therein.

30 The present invention further provides a negatively charged membrane comprising a porous substrate and a crosslinked coating. An embodiment of the process comprises:

(a) providing a porous substrate;

(b) contacting the substrate with a polysaccharide and a 35 polymerized composition comprising an unsaturated monomer having an anionic group, an N-(hydroxymethyl)- and/or N-(alkoxymethyl)- acrylamides, and an initiator;

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(c) curing the substrate obtained in (b) to obtain the negatively charged membrane; and

(d) optionally, extracting the membrane obtained in (c) to remove extractable residue therein.

5 The present invention further provides, in an embodiment, a negatively charged microporous membrane comprising a porous support and a crosslinked coating wherein the crosslinked coating is prepared from a polymerized composition comprising an unsaturated monomer having an anionic group, an N-(hydroxymethyl)- or N-(alkoxymethyl)-acrylamide, a nonionic hydrophilic acrylic monomer, and an initiator.

10 The present invention further provides, in another embodiment, a negatively charged microporous membrane comprising a porous substrate and a crosslinked coating prepared from a polysaccharide and a polymerized composition comprising an unsaturated monomer having an anionic group, an N-(hydroxymethyl)- or N-(alkoxymethyl)- acrylamide, and an initiator.

15 The present invention, in a further embodiment, provides a negatively charged microporous membrane comprising a porous substrate and a crosslinked coating prepared from a composition comprising an acrylic monomer having an anionic group, an N-(hydroxymethyl)- or N-(alkoxymethyl)- acrylamide, a nonionic hydrophilic acrylic monomer, and an initiator.

20 The present invention, in another embodiment, provides a negatively charged microporous membrane comprising a porous substrate and a crosslinked coating prepared from a polysaccharide and a polymerized composition comprising an acrylic monomer having an anionic group, an N-(hydroxymethyl)- or N-(alkoxymethyl)- acrylamide, and an initiator.

25 The present invention further provides a device e.g., a filter device, chromatography device, macromolecular transfer device, flow distributor arrangement, and/or a membrane module comprising one or more negatively charged membranes of the present invention. The device can be in any suitable form. For example, the device can include a filter element comprising the negatively charged membrane in a substantially

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planar or pleated form. In an embodiment, the element can have a hollow generally cylindrical form. If desired, the device can include the filter element in combination with upstream and/or downstream support or drainage layers. The 5 device can include a plurality of membranes, e.g., to provide a multilayered filter element, or stacked to provide a membrane module, such as a membrane module for use in membrane chromatography. Filter cartridges can be constructed by including a housing and endcaps to provide fluid seal as well 10 as at least one inlet and at least one outlet. The devices can be constructed to operate in crossflow or tangential flow mode as well as dead-end mode. Accordingly, the fluid to be treated can be passed, for example, tangentially to the membrane surface, or passed perpendicular to the membrane 15 surface.

For embodiments of the membrane which are in the form of a tube or fiber, or bundles of tubes or fibers, the membrane can be configured as modules, e.g., after potting their ends with an adhesive. For a description of illustrative 20 chromatographic devices, porous medium modules, and flow distributor arrangements, see U.S. Provisional Patent Application Nos. 60/121,667 and 60/121,701, both filed on February 25, 1999; U.S. Provisional Patent Application Nos. 60/168,738 and 60/168,750, both filed on December 6, 1999; and 25 International Applications filed on February 25, 2000 and entitled "Positively Charged Membrane" by Xiaosong Wu, Chung-Jen Hou, Jayesh Dharia, Peter Konstantin, and Yujing Yang; "Chromatography Devices and Flow Distributor Arrangements Used in Chromatography Devices" by Mark Hurwitz, Thomas Sorensen, 30 John Stempel, and Thomas Fendya; and "Chromatography Devices, Porous Medium Modules Used in Chromatography Devices and Methods for Making Porous Medium Modules" by Mark Hurwitz, Thomas Fendya, and Gary Bush. See also UK Patent Application GB 2 275 626 A.

35 The membrane of the present invention has one or more advantageous properties, including high water permeability dynamic protein binding capacity, and charge density. Thus,

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for example, the membrane preferably has a water flow rate above 5 mL/min/cm<sup>2</sup>, and preferably above 10 mL/min/cm<sup>2</sup>, e.g., from about 20 mL/min/cm<sup>2</sup> to about 160 mL/min/cm<sup>2</sup>, and preferably from about 25 mL/min/cm<sup>2</sup> to about 100 mL/min/cm<sup>2</sup> at 5 24 inch Hg. The membrane is robust and can withstand high treatment fluid flow rates. Thus, the membrane can be subjected to flow rates up to 10 cm/min, for example, from about 1 cm/min to 10 cm/min at 10 psi (68.9 kPa). The membrane has an open water bubble point of below about 70 psi 10 (482 kPa), e.g., from about 2.5 psi (9.39 kPa) to about 70 psi (482 kPa), and preferably from about 5 psi (34.47 kPa) to about 50 psi (344.7 kPa).

The membrane of the present invention has a high charge density. The charge density of the membrane can be measured 15 by methods known to those of ordinary skill in the art. For example, the charge density can be measured by the membrane's ability to bind a positively charged dye. Illustratively, the membrane has a Methylene Blue dye binding capacity of at least about 10 mL, e.g., from about 10 mL to about 1000 mL, and 20 preferably from about 100 mL to about 800 mL, when tested with a 10 ppm dye solution in water. Methylene Blue is a positively charged dye. The dye binding capacity is measured by filtering under a 24 inch Hg negative pressure, a 10 ppm by weight solution, pH 6.6, of Methylene Blue dye in a membrane 25 disc of 25 mm diameter, and monitoring the volume of the filtrate until a trace of the dye begins to appear in the filtrate.

The membrane of the present invention has a high specific protein binding capacity. The membrane has a lysozyme 30 specific binding capacity of above 10 mg/mL, e.g., from about 10 mg/mL to about 130 mg/mL and preferably from about 25 mg/mL to about 120 mg/mL. The specific binding capacity can be determined by the following illustrative method. A fluid containing a lysozyme protein in 10 mM MES buffer, pH 5.5, is 35 filtered by passing through a membrane at 1 cm/min and the concentration of the protein in the filtrate is measured as a function of time. The concentration of the protein can be

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determined spectrophotometrically, e.g., by measuring the

absorbance of the protein at 280 nm. A breakthrough curve such as the one shown in Fig. 1 can then be constructed with the x-axis depicting the time of the filtration experiment and the y-axis depicting the protein concentration in the  
5 filtrate. The membrane has high specific protein binding capacity and low non-specific or hydrophobic binding. The slope of the breakthrough curve obtained on the membrane is vertical or substantially vertical. This characteristic offers improved resolution and separation of proteins. The  
10 membrane also has high dynamic protein binding capacity.

An advantage of the membrane of the present invention is that proteins do not leak prior to breakthrough. Another advantage of the present invention is that the components of the membrane are carefully chosen so that the membrane is free  
15 or substantially free of grafts or covalent links between the coating and the substrate. The preparation of negatively charged membranes of the present invention involves a chemistry and procedure that is relatively simple and easy to practice.

20 The properties of the membranes of the present invention make them attractive for use in the detection, separation, and/or purification of biomolecules such as proteins, amino acids, nucleic acids, and viruses. Examples of nucleic acids include modified or unmodified, synthetic or natural RNA and  
25 DNA.

The membranes of the present invention find use in various applications such as filtration of fluids containing positively charged atoms, molecules, and particulates, and macromolecular transfer from electrophoresis gels such as the  
30 transfer of nucleic acids and proteins from electrophoresis gels to an immobilizing matrix. The membrane can find use in the separation or purification of components present in biological fluids. Thus, for example, the membrane can find use in the purification of human albumins from the serum, in  
35 the therapeutic fractionation of blood, and separation of the components in genetically engineered cell cultures or fermentation broths. The membrane can be used in the

purification of, for example, viral vaccines and gene therapy vectors such as adeno-associated viruses.

Accordingly, the present invention provides a process for treating a fluid containing biomolecules, the process

- 5 comprising placing the fluid in contact with the negatively charged membrane. The positively charged materials adsorbed on the membrane can be recovered by eluting with a suitable solvent eluant. The present invention further provides a process for selectively adsorbing one or more biomolecules
- 10 from fluid containing a mixture of biomolecules comprising contacting the fluid with the membrane under conditions favorable to the adsorption of selected biomolecules. The present invention further provides a process for selectively releasing one or more biomolecules from a membrane of the
- 15 present invention comprising contacting the membrane having adsorbed biomolecules with an eluant under conditions favorable to the release of the selected biomolecules. The present invention further provides a process for macromolecular transfer from an electrophoresis gel comprising
- 20 contacting a membrane of the present invention with the electrophoresis gel, and transferring the macromolecules from the gel to the membrane.

The negatively charged membrane of the present invention is particularly suitable for treating fluids containing biomolecules that have a positive surface charge for the given pH of the fluid. For example, lysozyme has an isoelectric point of 11.25, and it can be purified by using the negatively charged membrane of the present invention from a low salinity, for example 10mM MES, fluid that is pH 5.5. Proteins with different surface charges may also be separated using the membrane of the present invention, for example separating lysozyme from Cytochrome C.

Thus, a mixture of lysozyme and Cytochrome C can be separated as follows. 80 µl of a fluid containing 3 mg/ml lysozyme and 1 Cytochrome C can be placed on a chromatographic column or stack of 5 layers of a 25 mm diameter negatively charged membrane of the present invention. The column or

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stack can be eluted under a gradient - 7 ml from 10 mM MES buffer at a pH of 5.5 to 1M NaCl-10 mM MES buffer at a pH of 5.5. The flow rate can be 4 ml/min. Cytochrome C elutes first, followed by Lysozyme.

5 The following examples further illustrate the present invention but should not be construed in any way limiting the scope of the invention.

EXAMPLE 1

10 This Example illustrates a method of preparing a polymer composition for preparing an embodiment of the negatively charged membrane of the present invention.

15 2-Acrylamido-2-methyl-1-propanesulfonic acid, N-(isobutoxymethyl)acrylamide, and hydroxypropyl methacrylate were combined in a weight ratio of 8.0:2.5:1.5 in a methanol-water medium to obtain a polymerization solution having a solids content of 12% by weight. Ammonium persulfate was used as the initiator at 0.3% by weight of the solution. The polymerization was carried out for a period of about 10-15  
20 hours at ambient temperature (20-25°C). The resulting solution had a viscosity of 166 cps (0.166 Pa.s).

EXAMPLE 2

25 This Example illustrates a method for preparing an embodiment of the negatively charged membrane of the present invention. This Example further illustrates the properties of that embodiment.

30 A coating solution was prepared by mixing the polymerization solution described in Example 1 and a water solution of dextran, molecular weight 148 K, so that the resulting solution contains polymer and dextran in the weight ratio of 3:1.

35 A hydrophilic microporous polyethersulfone substrate having a pore size of about 0.8 µm was coated with the above coating solution. The coated substrate was cured in an oven at 100-110°C for 1 hour, followed by washing it in boiling DI

water for 1 hour. The resulting membrane was dried in an oven to obtain an embodiment of the present invention.

The membrane obtained above was tested for treatment of a solution containing lysozyme. The solution was contained

5 206.4 µg per ml of 10 mM MES buffer at pH 5.5. The treatment fluid flow rate was 4 ml/min. Two membrane discs of 25 mm diameter were stacked together. The breakthrough curve obtained is set forth in Fig. 1. The membrane had a lysozyme binding capacity of 97 mg/ml. The relatively flat curve  
10 obtained during the first 10 minutes of the treatment confirmed that the membrane did not leak. The nearly vertical slope indicates that the membrane was capable of providing high resolution.

15 EXAMPLE 3

This Example illustrates a method for preparing an embodiment of the negatively charged membrane of the present invention. This Example further illustrates the properties of that embodiment.

20 A coating solution was prepared by mixing the polymerization solution described in Example 1 and a water solution of dextran, molecular weight 148 K, so that the resulting solution contains polymer and dextran in the weight ratio of 4:1.

25 A hydrophilic microporous cellulose nitrate substrate having a pore size of about 0.8 µm was coated with the above coating solution. The coated substrate was cured in an oven at 100-110°C for 1 hour, followed by washing it in boiling DI water for 1 hour. The resulting membrane was dried in an oven  
30 to obtain an embodiment of the present invention.

The membrane obtained above was tested with a solution containing lysozyme. The solution was contained 201.3 µg per ml of 10 mM MES buffer at pH 5.5. The treatment fluid flow rate was 4 ml/min. Two membrane discs of 25 mm diameter were  
35 stacked together. The breakthrough curve obtained is set forth in Fig. 2. The membrane had a lysozyme binding capacity of 77 mg/ml. The relatively flat curve obtained during the

first 10 minutes of the treatment confirmed that the membrane did not leak. The nearly vertical slope indicates that the membrane was capable of providing high resolution.

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EXAMPLE 4

This Example illustrates a method for preparing another embodiment of the negatively charged membrane of the present invention. This Example further illustrates the properties of that embodiment.

10

2-Acrylamidoglycolic acid, 2-carboxyethyl acrylate, N-(isobutoxymethyl)acrylamide, N-(hydroxymethyl)-acrylamide, and hydroxypropyl acrylate were combined in a weight ratio of 5.0:5.0:3.0:1.5:1.5 in a methanol-water medium to obtain a polymerization solution having a solids content of 16% by weight. Ammonium persulfate was used as the initiator at 0.4% by weight of the solution. The polymerization was carried out for a period of about 16-24 hours at ambient temperature. The resulting solution had a viscosity of 116 cps (0.116 Pa.s). A coating solution was prepared by mixing the polymerization solution and a water solution of dextran, molecular weight 148 K, so that the resulting solution contained 4% polymer and 1.33% dextran by weight.

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A hydrophilic microporous polyethersulfone substrate having a pore size of about 0.8  $\mu\text{m}$  was coated with the above coating solution. The coated substrate was cured in an oven at 100-110°C for 1 hour, followed by washing it in boiling DI water for 1 hour. The resulting membrane was dried in an oven to obtain another embodiment of the present invention.

20

The membrane obtained above was tested with a solution containing lysozyme. The solution was contained 213.6  $\mu\text{g}$  per ml of 10 mM MES buffer at pH 5.5. The treatment fluid flow rate was 4 ml/min. Two membrane discs of 25 mm diameter were stacked together. The breakthrough curve obtained is set forth in Fig. 3. The membrane had a lysozyme binding capacity of 45 mg/ml. The relatively flat curve obtained during the first 10 minutes of the treatment confirmed that the membrane

did not leak. The nearly vertical slope indicates that the membrane was capable of providing high resolution.

All references cited herein, including patents, patent applications, and publications, are incorporated by reference  
5 in their entireties.

While this invention has been described with an emphasis upon several embodiments, it will be obvious to those of ordinary skill in the art that variations of the embodiments may be used and that it is intended that the invention may be  
10 practiced otherwise than as specifically described herein. Accordingly, this invention includes all modifications encompassed within the spirit and scope of the invention as defined by the following claims.

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## WHAT IS CLAIMED IS:

1. A negatively charged microporous membrane comprising a porous substrate and a crosslinked coating, wherein the crosslinked coating is prepared from a polymer comprising an unsaturated monomer having a negatively charged group, a hydrophilic non-ionic unsaturated monomer, and at least one or more N-(hydroxyalkyl)- or N-(alkoxyalkyl)- acrylamide.
2. The negatively charged microporous membrane of claim 1, wherein the hydrophilic non-ionic unsaturated monomer is an acrylic monomer.
3. The negatively charged microporous membrane of claim 1 or 2, wherein the N-(hydroxyalkyl)- or N-(alkoxyalkyl)- acrylamide includes an alkyl group of 4 carbon atoms or less.
4. The negatively charged microporous membrane of any of claims 1-3, wherein the crosslinked coating includes a hydroxyl-rich material.
5. The negatively charged microporous membrane of claim 4, wherein the hydroxyl-rich material is a polysaccharide.
6. The negatively charged microporous membrane of any of claims 1-5, wherein said negatively charged group is a sulfonic or carboxylic acid.
7. The negatively charged microporous membrane of claim 2, wherein said acrylic monomer is an acrylate or acrylamide.
8. The negatively charged microporous membrane of claim 7, wherein said acrylic monomer is an acrylamide.
9. The negatively charged microporous membrane of claim 8, wherein said acrylamide is an alkylacrylamide.

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10. The negatively charged microporous membrane of claim 9, wherein said acrylamide has a sulfonic acid group.

5 11. The negatively charged microporous membrane of claim 10, wherein said acrylamide is acrylamido-N-alkylsulfonic acid.

12. The negatively charged microporous membrane of claim 9, wherein said alkylacrylamide has a carboxylic acid group.

10

13. The negatively charged microporous membrane of claim 12, wherein said polymer includes a further acrylic monomer having a carboxylic acid group.

15

14. The negatively charged microporous membrane of claim 13, wherein said further acrylic monomer is an acrylate.

15. The negatively charged microporous membrane of claim 14, wherein said acrylate is  $\beta$ -carboxyethyl acrylate.

20

16. The negatively charged microporous membrane of claim 4, wherein said acrylic monomer is a hydroxyacrylic monomer.

25

17. The negatively charged microporous membrane of claim 16, wherein said hydroxyacrylic monomer is a hydroxyacrylamide or an hydroxyacrylate.

18. The negatively charged microporous membrane of any of claims 1-17, wherein said polymer includes an N-(alkoxymethyl)acrylamide.

30

19. The negatively charged microporous membrane of claim 5, wherein said polysaccharide is dextran.

35

20. The negatively charged microporous membrane of claim 1, wherein the polymer comprising an unsaturated monomer having a negatively

charged group, a hydrophilic non-ionic unsaturated monomer, and at least one or more N-(hydroxyalkyl)- or N-(alkoxyalkyl)- acrylamide is prepared by employing a free radical initiator.

5 21. The negatively charged microporous membrane of any of claims 1-  
20 having a dynamic protein binding capacity of about 25 mg/ml  
lysozyme or more.

10 22. A negatively charged microporous membrane comprising a porous substrate and a crosslinked coating comprising negatively charged groups and amide-amide and amide-ester crosslinks.

15 23. The negatively charged microporous membrane of any of claims 1-  
22, wherein said porous substrate comprises a substrate polymer.

20 24. The negatively charged microporous membrane of claim 23,  
wherein said substrate polymer is selected from the group  
consisting of polyaromatics, polysulfones, polyolefins,  
polystyrenes, polyamides, polyimides, cellulose acetates, cellulose  
nitrates, polycarbonates, polyesters, and fluoropolymers.

25 25. The negatively charged microporous membrane of claim 24,  
wherein said substrate polymer is a polysulfone.

26 26. The negatively charged microporous membrane of any of claims 1-  
25, wherein said porous substrate is hydrophilic.

27. A process for preparing a negatively charged microporous membrane comprising a porous substrate and a crosslinked coating having negatively charged groups, the process comprising:  
30 (a) providing a porous substrate;  
35 (b) contacting said substrate with a polymer comprising an unsaturated monomer having a negatively charged group, a hydrophilic non-ionic unsaturated monomer, and at least one or more of a N-(hydroxyalkyl)- or N-(alkoxyalkyl)- acrylamide;

(c) curing the substrate obtained in (b) to obtain the negatively charged microporous membrane; and

(d) optionally, extracting the membrane obtained in (c) to remove extractable residue therein.

5

28. A process for preparing a negatively charged microporous membrane comprising a porous substrate and a crosslinked coating having negatively charged groups, the process comprising:

(a) providing a porous substrate;

10 (b) contacting said substrate with a polysaccharide and a polymer comprising an unsaturated monomer having a negatively charged group and an N-(hydroxymethyl)- or N-(alkoxymethyl)- acrylamide;

15 (c) curing the substrate obtained in (b) to obtain the negatively charged microporous membrane; and

(d) optionally, extracting the membrane obtained in (c) to remove extractable residue therein.

20 29. The process of claim 27 or 28, wherein said negatively charged group is a sulfonic or carboxylic acid.

30. The process of any of claims 27-29, wherein said unsaturated monomer having a negatively charged group is an acrylic monomer having a sulfonic or carboxylic acid group.

25

31. The process of claim 30, wherein said acrylic monomer having a sulfonic or carboxylic acid group is an acrylate or an acrylamide.

32. The process of claim 27, wherein the substrate is contacted in  
30 (b) with said polymer and a hydroxyl-rich material.

33. The process of any of claims 27-32, wherein said porous substrate comprises a substrate polymer.

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charged group, a hydrophilic non-ionic unsaturated monomer, and at least one or more N-(hydroxyalkyl)- or N-(alkoxyalkyl)- acrylamide is prepared by employing a free radical initiator.

5 21. The negatively charged microporous membrane of any of claims 1-  
20 having a dynamic protein binding capacity of about 25 mg/ml  
lysozyme or more.

10 22. A negatively charged microporous membrane comprising a porous substrate and a crosslinked coating comprising negatively charged groups and amide-amide and amide-ester crosslinks.

15 23. The negatively charged microporous membrane of any of claims 1-  
22, wherein said porous substrate comprises a substrate polymer.

20 24. The negatively charged microporous membrane of claim 23,  
wherein said substrate polymer is selected from the group  
consisting of polyaromatics, polysulfones, polyolefins,  
polystyrenes, polyamides, polyimides, cellulose acetates, cellulose  
nitrates, polycarbonates, polyesters, and fluoropolymers.

25 25. The negatively charged microporous membrane of claim 24,  
wherein said substrate polymer is a polysulfone.

26 26. The negatively charged microporous membrane of any of claims 1-  
25, wherein said porous substrate is hydrophilic.

27. A process for preparing a negatively charged microporous  
membrane comprising a porous substrate and a crosslinked coating  
30 having negatively charged groups, the process comprising:

(a) providing a porous substrate;

(b) contacting said substrate with a polymer comprising an  
unsaturated monomer having a negatively charged group, a  
hydrophilic non-ionic unsaturated monomer, and at least one or more  
35 of a N-(hydroxyalkyl)- or N-(alkoxyalkyl)- acrylamide;

(c) curing the substrate obtained in (b) to obtain the negatively charged microporous membrane; and

(d) optionally, extracting the membrane obtained in (c) to remove extractable residue therein.

5

28. A process for preparing a negatively charged microporous membrane comprising a porous substrate and a crosslinked coating having negatively charged groups, the process comprising:

(a) providing a porous substrate;

10 (b) contacting said substrate with a polysaccharide and a polymer comprising an unsaturated monomer having a negatively charged group and an N-(hydroxymethyl)- or N-(alkoxymethyl)- acrylamide;

15 (c) curing the substrate obtained in (b) to obtain the negatively charged microporous membrane; and

(d) optionally, extracting the membrane obtained in (c) to remove extractable residue therein.

20 29. The process of claim 27 or 28, wherein said negatively charged group is a sulfonic or carboxylic acid.

30. The process of any of claims 27-29, wherein said unsaturated monomer having a negatively charged group is an acrylic monomer having a sulfonic or carboxylic acid group.

25

31. The process of claim 30, wherein said acrylic monomer having a sulfonic or carboxylic acid group is an acrylate or an acrylamide.

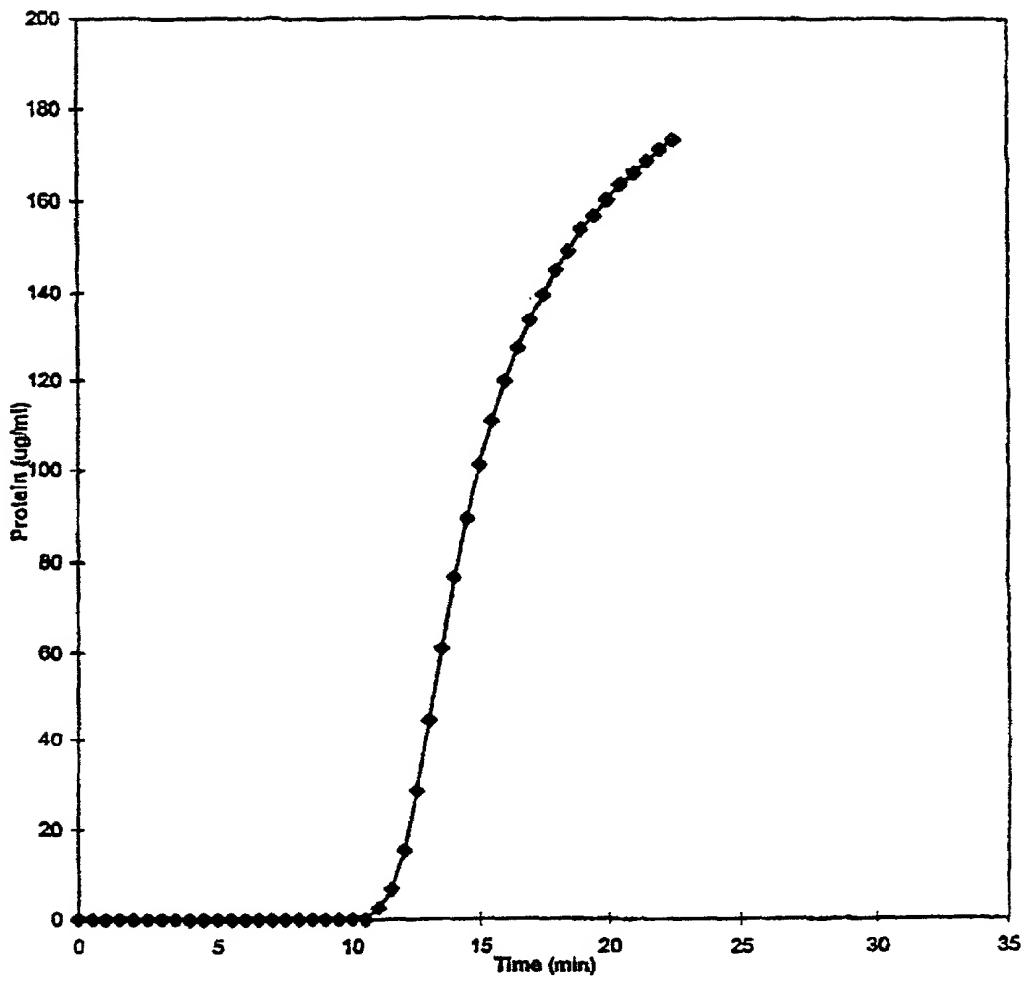
30 32. The process of claim 27, wherein the substrate is contacted in (b) with said polymer and a hydroxyl-rich material.

33. The process of any of claims 27-32, wherein said porous substrate comprises a substrate polymer.

34. The negatively charged microporous membrane prepared by the process of any of claims 27-33.
35. A device comprising the negatively charged microporous membrane of any of claims 1-26 and 34.
36. A process for separating positively charged material from a fluid, said process comprising placing said fluid in contact with the negatively charged microporous membrane of any of claims 1-26 and 34 so as to adsorb the positively charged material to said membrane.
37. The process of claim 36, wherein said positively charged material is a biomolecule.
38. A process for transferring biomolecules from an electrophoresis gel comprising contacting said electrophoresis gel with a membrane of any of claims 1-26 and 34 and transferring the biomolecules to the membrane.
39. The process of claim 38, wherein said biomolecule is selected from the group consisting of proteins, polypeptides, amino acids, and nucleic acids, and combinations thereof.
40. The process of claim 38 or 39, further including recovering the positively charged material adsorbed on the membrane.

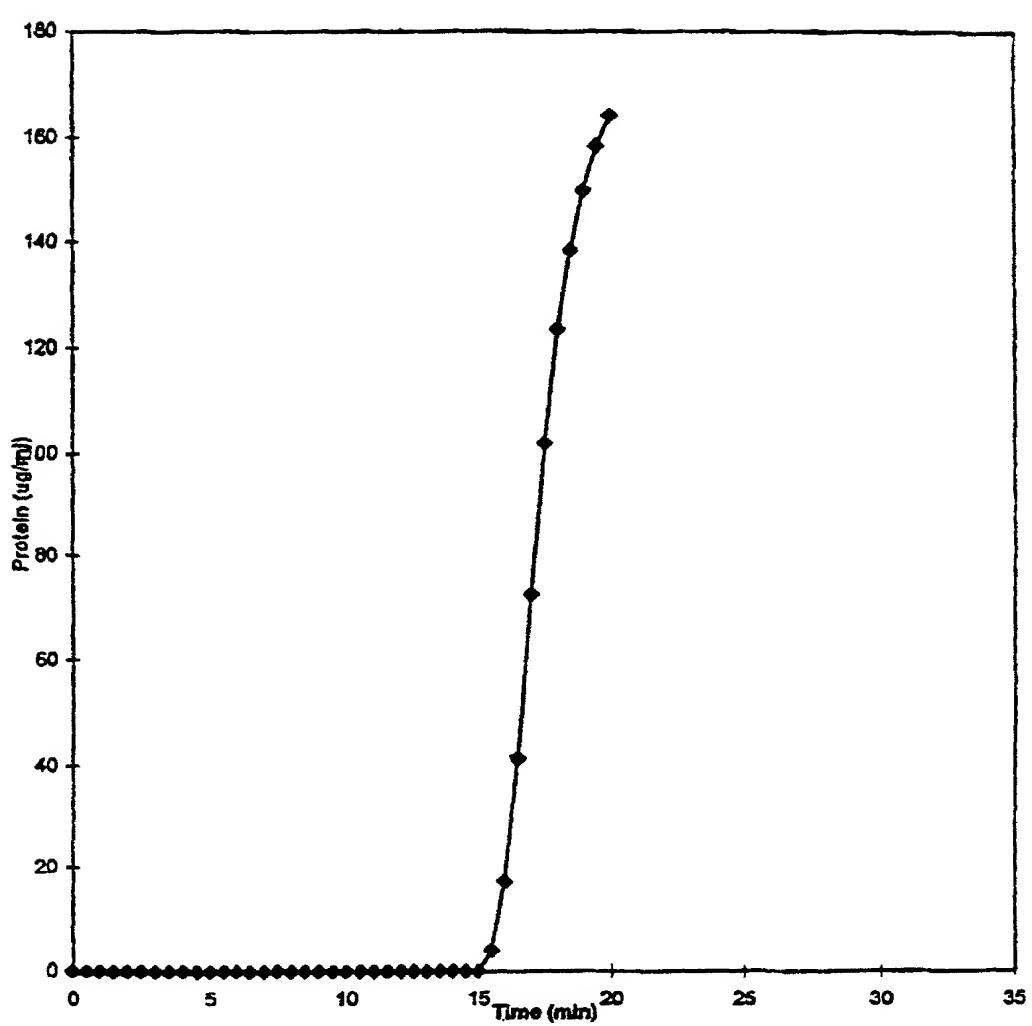
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Figure 1



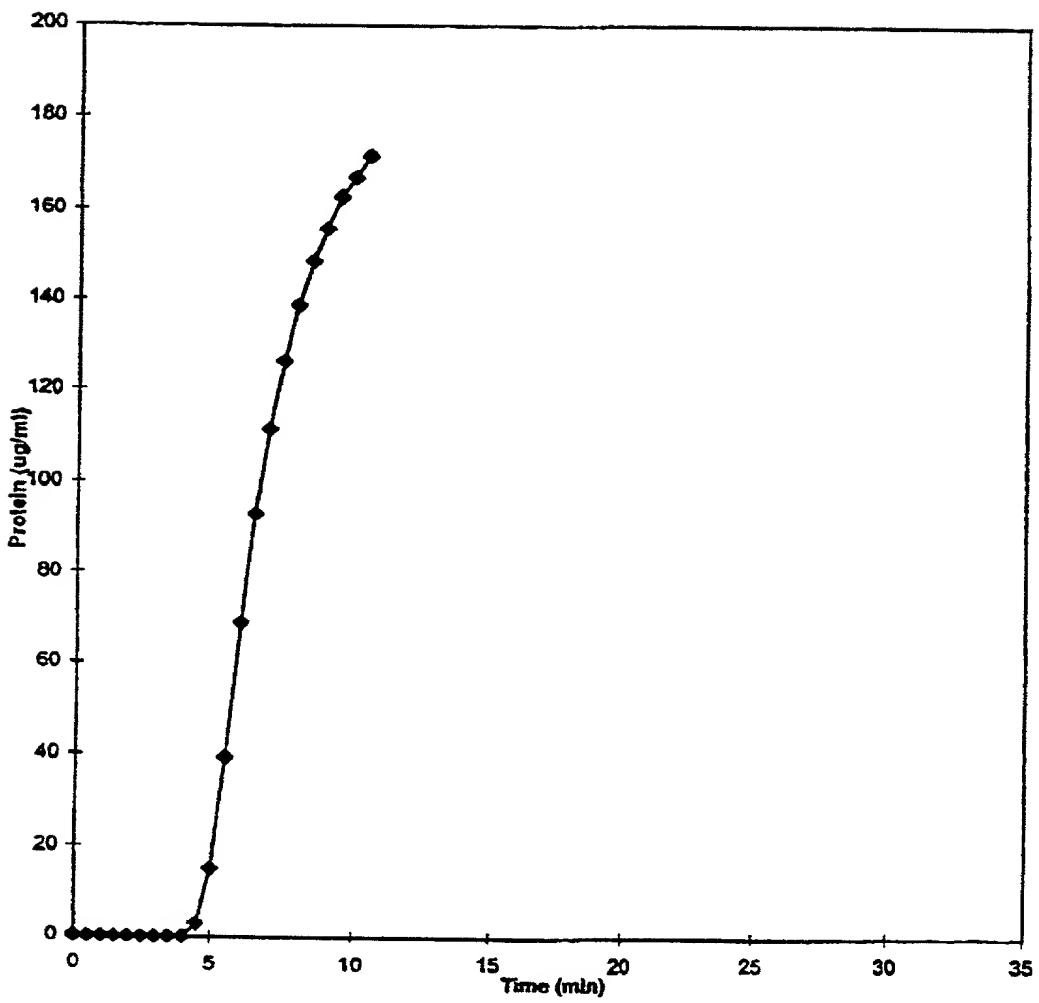
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Figure 2



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Figure 3



**PATENT**  
Attorney Docket No. 440508

**COMBINED DECLARATION AND POWER OF ATTORNEY**

As a below named inventor, I hereby declare that:

This declaration is of the following type:

- original    design    supplemental  
 national stage of PCT  
 divisional    continuation    continuation-in-part

My residence, post office address, and citizenship are as stated below next to my name. I believe I am the original, first, and sole inventor (*if only one name is listed below*) or an original, first, and joint inventor (*if plural names are listed below*) of the subject matter which is claimed and for which a patent is sought on the invention entitled:

**NEGATIVELY CHARGED MEMBRANE**

the specification of which:

- is attached hereto.  
 was filed on \_\_\_\_\_ as Application No. \_\_\_\_\_ and was amended on \_\_\_\_\_ (*if applicable*).  
 was filed by Express Mail No. \_\_\_\_\_ as Application No. *not known yet*, and was amended on \_\_\_\_\_ (*if applicable*).  
 was filed on February 25, 2000 as PCT International Application No. PCT/US00/04745 and was amended on August 24, 2001 (*if any*).

I state that I have reviewed and understand the contents of the specification identified above, including the claim(s), as amended by any amendment referred to above.

I acknowledge the duty to disclose information that is material to the patentability of the application identified above in accordance with 37 CFR 1.56.

I claim foreign priority benefits under 35 USC 119(a)-(d) or 365(b) of any foreign application(s) for patent or inventor's certificate or 365(a) of any PCT international application(s) designating at least one country other than the United States of America listed below and have also identified below any foreign application(s) for patent, utility model, design registration, or inventor's certificate or any PCT international application(s) designating at least one country other than the United States of America filed by me on the same subject matter and having a filing date before that of the application(s) from which the benefit of priority is claimed.

PRIOR FOREIGN PATENT, UTILITY MODEL, AND DESIGN REGISTRATION APPLICATIONS						
COUNTRY	PRIOR FOREIGN APPLICATION NO.	DATE OF FILING (day,month,year)	PRIORITY CLAIMED			
				YES		NO
				YES		NO
				YES		NO

I claim the benefit pursuant to 35 USC 119(e) of the following United States provisional patent application(s):

PRIOR U.S. PROVISIONAL PATENT APPLICATIONS, BENEFIT CLAIMED UNDER 35 USC 119(e)	
APPLICATION NO.	DATE OF FILING (day,month,year)
60/121,668	25 February 1999

I claim the benefit pursuant to 35 USC 120 of any United States patent application(s) or PCT international application(s) designating the United States of America listed below and, insofar as the subject matter of each of the claims of this patent application is not disclosed in the prior patent application(s) in the manner provided by the first paragraph of 35 USC 112, I acknowledge the duty to disclose material information as defined in 37 CFR 1.56 effective between the filing date of the prior patent application(s) and the national or PCT international filing date of this patent application.

PRIOR U.S. PATENT APPLICATIONS OR PCT INTERNATIONAL APPLICATIONS DESIGNATING THE U.S., BENEFIT CLAIMED UNDER 35 USC 120				
U.S. PATENT APPLICATIONS		Status ( <i>check one</i> )		
U.S. APPLICATION NO.	U.S. FILING DATE	PATENTED	PENDING	ABANDONED
1.				
2.				
3.				
PCT APPLICATIONS DESIGNATING THE U.S.		Status ( <i>check one</i> )		
PCT APPLICATION NO.	PCT FILING DATE (day,month,year)	U.S. APPLICATION NOS. ASSIGNED (if any)	PATENTED	PENDING
4.				
5.				
6.				

DETAILS OF FOREIGN APPLICATIONS FROM WHICH PRIORITY CLAIMED UNDER 35 USC 119 FOR ABOVE LISTED U.S./PCT APPLICATIONS				
ABOVE APPLICATION. NO.	COUNTRY	APPLICATION NO.	DATE OF FILING (day,month,year)	DATE OF ISSUE (day,month,year)
1.				
2.				
3.				
4.				
5.				
6.				

As a named inventor, I appoint the following registered practitioner(s) to prosecute this application and transact all business in the Patent and Trademark Office connected therewith: Customer Number 23548.



I further direct that correspondence concerning this application be directed to Customer Number 23548.



I declare that all statements made herein of my own knowledge are true, that all statements made on information and belief are believed to be true, that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code, and that such willful false statements may jeopardize the validity of the application or any patent issued thereon.

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Full name of second joint inventor: Peter KONSTANTIN

Inventor's signature \_\_\_\_\_

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As a named inventor, I appoint the following registered practitioner(s) to prosecute this application and transact all business in the Patent and Trademark Office connected therewith: Customer Number 23548.



I further direct that correspondence concerning this application be directed to Customer Number 23548.



I declare that all statements made herein of my own knowledge are true, that all statements made on information and belief are believed to be true, that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code, and that such willful false statements may jeopardize the validity of the application or any patent issued thereon.

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DEC-00-2001 10:44

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